Production ecology of the amphipod Hyalella azteca (Saussure) in a Northern Ontario lake
by


A thesis
submitted to the Department of Biology in partial fulfillment of the requirements for the degree of Master of Science

Lakehead University
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## ABSTRACT

Annual production of the amphipod Hyalella azteca was estimated in the upper littoral zone of Dock Lake, Ontario, over a 2 year period (1980-1981). Population density (negligible below 1 meter) differed among the three habitats sampled. Cohort production by the size frequency method was $1.39 \mathrm{~g} / \mathrm{m}^{2}$ (dry weight), mean annual biomass was $0.36 \mathrm{~g} / \mathrm{m}^{2}$, and the $\mathrm{P} / \overline{\mathrm{B}}$ ratio was 3.9 for the 1980 year class. Six other methods for estimating production were applied to this cohort, providing comparable estimates ranging from 1.21 (instantaneous growth) to 1.65 (cohort G) g/m². Size frequency estimates of annual production for 1980 and 1981 were 1.24 and $1.40 \mathrm{~g} / \mathrm{m}^{2}$ respectively. The annual $P / \bar{B}$ ratios of 3.9 and 4.0 for these two years agree with the cohort value. A comparison of production estimates for Hyalella azteca in the literature shows a gradient of production correlating with habitat temperature regime.

This thesis is an original composition, based on research carried out by the author, and has not been previously submitted for credit toward any degree or diploma. Where the work of others has been included, it has been so acknowledged and appropriately cited.


August 16, 1982.

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Figure 1: Depth contour map of Dock Lake, Ontario. Production sampling stations consisted of three 5 meter lengths of shoreline (rectangles). Preliminary survey transects ran north-south, with two single-station transects at the east and west ends of the lake. Depth distribution survey transects (not shown) were taken perpendicular to the shore at each production site.

Figure 2: Dock Lake shore temperatures for the ice-free seasons of 1980 and 1981 on the three sample sites; Carex, Typha and Gravel shores.

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Figure 6: Growth curve for Hyalella azteca, from instars 1 through 10. Molt rates for juveniles, and incubation rates (corresponding to adult female molt rates) are taken from Cooper (1965). Weight is weight at average length for each instar, calculated from the length-weight regression.(32)

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Figure 7: Interaction of climate with production (circles) and biomass (triangles) in $\mathrm{g} / \mathrm{m}^{2}$ dry weight of Hyalella azteca in four temperate zone lakes. Number of days above $20^{\circ} \mathrm{C}$ reflects optimal growth conditions; number of days above $10^{\circ} \mathrm{C}$ is an interaction of climate and lake morphometry.

Production is 'the means by which energy is made available for transmission from one trophic level to the next" (Waters, 1977). Studies of the production of detritivores thus form a link in the understanding of an energy pathway of major significance in aquatic ecosystem metabolism (Brinkhurst, 1974; Wetzel, 1975).

The common amphipod Hyalella azteca is a widely distributed freshwater detritivore. Often a major component of the benthos (see Anderson and Hooper, 1956), it is preyed upon by such fish as Stizostedion vitreum, Perca flavescens, Oncorhynchus nerka, and Salmo gairdneri (see Fedoruk, 1966; Cooper, 1965; Mathias, 1971), as well as invertebrate predators (Jackson, 1912).

Turnover, or $P / \bar{B}$ ratio values approximate 4 to 6 for most benthic invertebrates, and may thus be useful in estimating production (Waters, 1979). Calculation of the $P / \bar{B}$ ratio for $\underline{H}$. azteca in different habitats and climates may help elucidate the interaction between environment and population dynamics; hence providing a way to develop predictive equations for estimating benthic production of lakes, as well as understanding the role of detritivores in aquatic ecosystems.

By comparing the production and turnover ratio of $\underline{H}$. azteca in Northwestern Ontario with other available production estimates
(see Cooper, 1965; Mathias, 1971), the modifying effects of energy availability on life cycle and population dynamics of this important benthic species can be measured.

Life history of Hyalella azteca
H. azteca lives on or under the surface of detrital debris, and is particularly common among beds of emergent macrophytes. It feeds on dead and living plant material, especially filamentous algae, and epiphytic growth (Jackson, 1912; Embody, 1912; Cooper, 1965), although it will also consume fine detritus (Hargrave, 1970).

This amphipod is ubiquitously distributed in all North American permanent fresh water that reaches a monthly mean temperature of more than $10^{\circ} \mathrm{C}$ (Bousfield, 1958). It has been reported from such northerly sites as Great Slave Lake (Moore, 1980), White Horse, Yukon, and lakes north of Rampart House, Alaska (Pearse, 1913). It is also common in Central and South America to Tierra del Fuego (Bulycheva, 1957).
H. azteca inhabits the littoral zone of lakes, but can be found to a depth of 10 meters (Bousfield, 1973). Cooper (1965) found it homogenously distributed above 1.75 m . in Sugarloaf Lake, Michigan. In Marion Lake, British Columbia, Mathias (197l) found it to be most numerous (up to $5000 / \mathrm{m}^{2}$ ) in very shallow water, but decreasing 10 -fold from 0.5 m . to 3 m . Mean depth of Marion and

Sugarloaf Lakes are 2.4 m., and approximately 1.2 m., respectively.

The life cycle of this amphipod is univoltine. An overwintering population begins to breed in early to late May, or early June, depending on water temperature. The young grow rapidly. Some broods may attain sexual maturity in their first summer (Gaylor, 1922; Cooper, 1965; Biette, 1969), however, Mathias (1971) found that in Marion Lake, which rarely goes above $20^{\circ} \mathrm{C}$, the amphipods did not mature until their second summer. Reproduction is continuous during the summer for as long as temperature permits, resulting in overlap of year classes and broods within a year class. Sex recognition is primarily by touch (Holmes, 1903), and amplexus precedes mating. The male carries the female until she molts; immediately after which sperm is deposited in the female's brood pouch, or marsupium. Eggs are subsequently passed from the ovary to the marsupium, where they are fertilized (Jackson, 1912; Geisler, 1944). Clutch size varies from 4 to 40 , being a function of female body size (Mathias, 1971). Males can be distinguished from females by the enlarged second gnathopod, presence of genital papillae ventrally on peraeon 7, and absence of coxal oostegites (Bousfield, 1973). Breeding condition in females can be estimated by the appearance of the oostegites, even when there are no eggs in the marsupium. Non-breeding is indicated by poorly developed oostegites which lack a fringe of long bristles (Geisler, 1944).

The eggs develop and hatch within the marsupium. The young remain within the pouch for one to three days, then work their way out (Geisler, 1944), or are released when the female molts (Gaylor, 1922). Cooper (1965) has shown that each juvenile instar has a characteristic mean number of antennal segments; however, adults are not as consistent. Instars 1 through 7 constitute the immature stages, instar 8 and above are considered adults. Instar 7 has nine or ten segments on the first antenna, with a maximum of twenty-two on the first and second antennae on one side of the head (Cooper, 1965; Geisler, 1944).

Temperature and photoperiod are critical factors regulating growth and reproduction. Cooper (1965) and Bovee (1950) demonstrated that the molt rate, incubation rate and age to maturity are a function of temperature; that growth is negligible at $10^{\circ} \mathrm{C}$, and optimal ranges for growth and reproduction are 20 to $25^{\circ} \mathrm{C}$.

Photoperiod determines reproductive state (active or resting), and temperature influences the rate of change from one state to the other. Breeding is initiated by a light regime of 12L-12D when temperature rises above $16^{\circ} \mathrm{C}$ (breeding can be induced at $10^{\circ} \mathrm{C}$ if this photoperiod is maintained for 10 weeks), similarly a return to this photoperiod from a longer day will halt breeding if the temperature drops (de March, 1977).

Size at maturity is dependent on the temperature experienced by the egg and embryo. Temperatures of 10 to $15^{\circ} \mathrm{C}$ produce large adults with delayed reproductive development, 20 to $23^{\circ} \mathrm{C}$ produces
small adults, and $18^{\circ} \mathrm{C}$ produces intermediate sized animals (de March, 1978).

## Study area

Dock Lake (Figure 1) is one of four marl lakes situated 10.5 km . west of Lake Superior, within Thunder Bay, Ontario (lat. $48^{\circ} 28^{\prime}$, long. $89^{\circ} 28^{\prime}$ ). It is 1.2 hectares in area, with a mean depth of 4.5.m. (Momot, 1973). Emergent macrophytes of the shoreline are dominated by Carex aquatilis and Typha latifolia. Submergent vegetation includes thick beds of Potamogeton spp. and Chara sp. Momot (1978) lists fish species, vegetation, and describes the limnology of this lake.

Beach temperatures during the ice-free seasons of 1980 and 1981 are shown in Figure 2. During the cool spring in 1981, the lake remained below the $20^{\circ} \mathrm{C}$ range until a much later date than in 1980.

The lake is stratified for most of the year. Conditions in the epilimnion vary with weather, but the hypolimnion is very stable. The thermocline is at 3 m ., below which anoxia exists until turnover in the late fall. Winter conditions vary; in 1980 the lake was anoxic below 2 m. , in 1981 it was oxic.

During the summer, conductivity readings in the epilimnion range from 280 to 310 micromhos. The hypolimnion exhibits a gradient of 300 micromhos at three meters, to 400 micromhos at the bottom. The water column as a whole drops briefly to 200 micromhos during turnover.


FIGURE 1: Depth contour map of Dock Lake, Ontario. Production sampling stations consisted of three 5 meter lengths of shoreline (rectangles). Preliminary survey transects ran north-south, with two single-station transects at the east and west ends of the lake. Depth distribution survey transects (not shown) were taken perpendicular to the shore at each production site.



FIGURE 2: Dock Lake shore temperatures for the ice-free seasons of 1980 and 1981 on the three sample sites; Carex, Typha and Gravel shores.

Secchi depth remains at 2 m . from breakup to midAugust, but ranges from 1.0 to 1.5 m . during fall.

Dock Lake exhibits a blend of marl, bog, and eutrophic characteristics. Wetzel (1975) describes the potential for rapid change in small marl lake ontogeny inherent in the development of littoral flora, especially Sphagnum. Approximately $50 \%$ of Dock Lake's shoreline is composed of floating vegetation mats containing this moss. The evidence suggests that Dock Lake is entering senescence, with the "pseudo-oligotrophy" (Momot, pers. comm.) imparted by marl chemistry giving way to eutrophic characteristics; eventually reaching a bog state.

## MATERIALS AND METHODS

## Limnology

Maximum/minimum thermometers were installed at three stations, each representative of a shore type (see Figure l). The thermometers were read on a weekly schedule during the icefree seasons of 1980 and 1981. Mid-lake profiles of temperature, oxygen and conductivity were taken using YSI Company meters, and Secchi depth recorded, weekly, for the 1980 season.

## Preliminary Survey

A survey was performed in early May, 1980, to establish the distribution and density of $H$. azteca in Dock Lake. Transect sampling was used to test for variability in depth distribution. The survey consisted of four transects plus two single shore stations (see Figure l), for a total of 26 samples. Two replicates were taken per station, as Cuff and Coleman (1979) found that survey precision was increased by increasing the number of stations sampled at the expense of the number of replicates per station.

Offshore stations were sampled with a small Ponar grab ( $259 \mathrm{~cm}^{2}$ ). Depths 0.5 m . or less were sampled using a cylindrical box sampler (CBS) (Wilding, 1940; Hynes, 1970, p 238) of $919 \mathrm{~cm}^{2}$ area. Samples were collected from the CBS using a fine mesh (0.25 micrometer) hand net (Hynes, 1971, pp 69-70; Deacon, 1979). To reduce variation in sampling effort a standardized method was adopted; of heavy vegetation removal, counted sweeps with the net,
and a timed break between sets of sweeps. As $\underline{H}$. azteca swims rapidly about when disturbed, this break proved a useful indicator of the preceding sweeps' effectiveness. Tests showed that the CBS was 90 to $100 \%$ effective in capturing amphipods on a gravel substrate; however, this value could be much lower in heavy vegetation.

Unpreserved field samples were transported to the laboratory in gallon jars, and washed in a \#60 Tyler sieve (0.25 micrometer mesh), to remove silt. Cooper (1965) found this mesh size to retain the smallest size classes. Subsequently, measurements of head width of newly hatched individuals supported the use of the 0.25 micrometer mesh sieve (see Jonasson, 1955). Head width was greater than 0.25 micrometers for young within marsupiae, whereas free-swimming young are larger. The samples were stored in $10 \%$ formalin. Howmiller (1972), and Mills, Pitman and Munroe (1982) found that benthic organisms undergo the least weight loss in formalin.

Sugar flotation (Anderson, 1959) can be 90 to $99 \%$ effective in extraction of benthos from samples (Gerking, 1962; Cooper, 1965). In this study, the CBS samples contained large amounts of plant matter. As a result, the sugar flotation became a mere concentrating step, because the amphipods could not be skimmed from the surface of the sugar solution without also acquiring considerable plant material. This skimmed material was then hand-picked under a dissecting microscope to extract the amphipods.

Extracted amphipods were sorted into males, females and juveniles (instars 7 and younger), and counted. All samples in this study were treated similarly.

To check the preliminary survey, and test for seasonal habitat changes three additional depth distribution surveys were performed, in July, October and March. The former two consisted of one transect off each production station, with five stations per transect in July, and four in September. Due to adverse ice conditions, only three stations were sampled in March. In each survey, three replicates were taken at each station with the Ponar grab.

Taylor's Power Law was used to transform the density data (Elliot, 1977). The appropriate transformation is $x^{P}$, where $x$ is the number of animals in the sample. Values of $P$ obtained were 0.20 for the distributional data, and 0.25 for the production samples (see below). This agrees with the general transformation of $x^{0.25}$ calculated by Downing (1979). The distributional data were analyzed for trends in depth, transect, location and seasonality.

## Length-Weight

Length-weight samples were collected in April, twice each in June and August, and in October, 1981, using a fine mesh D-net. Once extracted and sorted, the amphipods were stored in formalin except for a portion of the late June sample in which the animals were heat-killed, and processed immediately, to obtain fresh weight
measurements.
Head length was measured in profile as the arc between the tip of the rostrum and the dorsal margin of the head capsule, to the nearest hundredth of a millimeter using an ocular micrometer. Segment number counts for antenna 1 , and first and second antennae were recorded. Total length measurements involved a mechanism similar to that described by Anderson and Hooper (1956). Slots of varying width were cut in a plexiglass plate, and sections of fine ruler ( 1 division $=0.42 \mathrm{~mm}$.) were attached beside them. Each individual was inserted into the appropriate width slot, dorsal surface uppermost, and straightened by pressing gently on its back with fine forceps. Length was measured from the anterior of the rostrum to the tip of the telson.

Individuals were recorded as male, female, or juvenile. Eggs and/or young were removed from marsupiae by pressing them out through the interlocking hairs of the oostegites for counting.

Once measured, the animals were placed on weighing papers. Adults were treated individually and in groups of two to four animals of identical morphometrics and sex. Juveniles were similarly placed into groups of identical animals, in which the number of individuals per group was increased as size decreased; up to 100 or more for eggs and young from brood pouches. After drying for three days at $60^{\circ} \mathrm{C}$, the amphipods were weighed on a five place Mettler electronic balance.

A total of 2861 individuals were measured for length-weight
analysis. With grouping, these animals gave 674 weight data. Nine measurements of egg weight were obtained, involving 920 eggs. The weight data were transformed, and analyzed for differences between males and females, preserved and unpreserved samples, and season. Linear regression was used to obtain a length-weight equation.

## Production

Restricted distributions and preferred habitats are important factors to consider in designing sampling programs appropriate to the life history of the organism under study (Malley and Reynolds, 1979; Hall, Waters and Cook, 1980). Because the largest number of $\underline{H}$. azteca occurred in less than 0.5 m . of water, an extreme form of stratified random sampling, employing the cylindrical box sampler, was used to obtain production samples. Three strata of habitat-type were chosen (see Green, 1979; Hall, Waters and Cook, 1980). The three main habitat types in Dock Lake are beds of Carex aquatilis, stands of Typha latifolia, and gravel beach containing sparse growths of Equisetum sp. and Chara sp. Five meter sections of shoreline served as stations. Samples were taken randomly within each station as in Green and Hobson (1970); within the limitations of the sampler, and the irregularities of the Carex and Typha vegetation mats.

Using the mean density of $\underline{H}$. azteca obtained from the preliminary survey, and the area of the $C B S\left(919 \mathrm{~cm}^{2}\right)$, the optimum
replicate number of three was obtained from Table 6 of Downing (1979). Therefore, three replicates were taken per station on each sampling date.

Waters (1969) recommends sampling schedules tailored to the growth type of the study organism. Cooper (1965) indicated that $\underline{H}$. azteca juveniles can molt every 5 days at $20^{\circ} \mathrm{C}$, therefore the 1980 sampling schedule consisted of samples at five day intervals during the optimal season, and weekly to bi-weekly samples when water temperatures fell below this threshold. In 1981, a regular bi-weekly sampling schedule was adopted. Because the CBS did not function on ice-bound shores, samples could only be taken during the ice-free season.

Amphipod densities obtained in the production samples were tested for differences among stations on a sample by sample and total basis, and for trends in depth distribution within the range of the samples.

Once counted, the amphipods obtained from each replicate sample were stored in separate "adult" and "juvenile" vials. The three replicates of each sample were pooled, because size frequencies did not significantly differ among replicates. If the contents numbered more than approximately 100 individuals, adult and juvenile vials were subsampled using a plankton splitter, or a gridded petri dish and a table of random numbers. Cumulative subsampling revealed that a subsample of 75 to 100 animals accurately established the size frequencies of individuals within each vial.

Matching size classes to instars increases the accuracy of
production estimates which use the Hynes method (Hudson and Swanson, 1972; Resh, 1979). Therefore a graph of all measured animals of the 1980 data was examined for peaks indicative of mean instar sizes. Using mean instar antennal segment counts from Cooper (1965) as a criterion, the average head length and total length of instars 1 through 9 were calculated, compared to the graph peaks, and later checked against the length-weight data. From a total of 55,846 animals captured, 13,411 were measured for size frequency analysis.

After appropriate size classes were established, the measurement data were grouped into percentage frequencies for each size class. These were applied to the counts of juveniles and adults for each replicate, and to the sample means. Size frequency histograms of percentages and numbers for each sample were used in conjunction with lake temperature data (Figure 2), and the temperature dependent molt rates from Cooper (1965) to interpret life history.

Comparison of modal progressions of hatch peaks established that Cooper's molt data adequately fit the Dock Lake population. A modification of instar analysis (Cooper, 1965) was used to predict the expected location of hatch peaks in each sample, using mean temperatures over sample intervals and molt rates at those temperatures. The duration (in days) between samples was divided by the temperature-specific duration of instars (for juveniles) and incubation (an approximation of adult female molt rate, obtained from Cooper, 1965). The result is the potential number of instars (size classes)
the individuals of a hatch can grow through or into, during each sample interval. These calculations served to supplement the size frequency histograms, which were difficult to interpret. This information, together with temperature dependent size at maturity (de March, 1978) helped establish voltinism (see Table 3).

The numerical size frequency data, sampling schedule, and mean weight at mid length of each size class from the lengthweight analysis were utilized in a computer program for estimation of production by the size frequency method (Krueger and Martin, 1980). Production for the 1980 and 1981 seasons, and for the 1980 year class (cohort) was estimated. A cohort is herein considered as "a group of individuals all hatched over the entire summer" (Hudson and Swanson, 1972). Separate production estimates for each of the three shores sampled were combined into a weighted mean, using the relative proportions of each habitat. Carex comprised $34 \%$, Typha $42 \%$, and Gravel $24 \%$ of the total shoreline perimeter.

The size frequency method (Hynes and Coleman, 1968;
Hamilton, 1969), with the application of the cohort production interval voltinism correction (Benke, 1979), is comparable to other methods in accuracy. The size frequency method was chosen as the standard for this study.

For comparison, production was also estimated for the 1980 year class using the cohort $G$ (Mathews, 1970), daily G (Hall, Waters and Cook, 1980; Waters, 1981), instantaneous growth (Ricker, 1946; Allen, 1949), Allen Curve (Allen, 1951), and removal summation
(Anderson and Hooper, 1956; Teal, 1957) methods, as outlined in Waters and Crawford (1973), and Wetzel and Likens (1979). Waters (1977) fully discusses the application of these methods. Year class production and biomass values were multiplied by $365 / 412$ to obtain the annual values, because the 1980 year class was present in the samples for 412 days. Annual values are therefore $88.6 \%$ of year class values.

All statistical tests were performed at the $95 \%$ level of significance.

## Distribution

In Dock Lake, H. azteca is restricted to shallow water, with no individuals found below 1.5 meters (Figure 3).

No seasonal changes occur in depth distribution. The Kruskal Wallis test and parametric analysis of variance showed no significant difference in numbers of amphipods per m${ }^{2}$ (transformed and untransformed data) at equal depths among transects, within replicates, or among samples. Significant differences in numbers of $\underline{H}$. azteca existed between depths in each sample. The 0 to 1 meter depth division significantly differed from all deeper samples within each transect, and in the pooled data. A finer depth scale was used for analysis of the May and July samples. The 0 to 0.5 m range differed significantly from all deeper sets.

Separate production samples from each shore type were analyzed for differences in depth distribution, and also as a pooled set for each year. Depths of the production samples ranged from 0.08 to 0.5 meters, as the CBS did not function in less than 5 cm of water. The pooled sets for both years show a similar trend, with few animals found in deeper samples, while many occurred in the shallower samples; however the individual


FIGURE 3: Distribution of Hyalella azteca with depth in Dock Lake; including all preliminary survey and depth distribution samples (total number of samples was 139). Zero values represent multiple samples at all depths greater than 1 meter.
shores showed no significant difference among numbers of animals at different depths. Using a 0.2 m increment scale of depth, the following results were obtained for total numbers of animals, and numbers of adults in each depth increment: parametric analysis of variance showed no significant difference, the Kruskal Wallis test gave significance, and three ranges tests (Duncan's Multiple Range, Modified LSD, and Scheffe's tests) indicated that the Carex shore differed from the Typha and Gravel shores.

Kruskal Wallis tests for differences among numbers of animals on the three shores showed significance (pooled 1980 and 1981 data). Parametric ANOVA indicated significant differences in total numbers of animals among the three shores for 1980 but not 1981. Numbers of adults and juveniles in 1980, and adults only in 1981 were significantly different among the three shores. In all cases the ranges tests showed that the Carex samples were different from those of Typha and Gravel (see Figure 5).

On a sample by sample basis, the 1980 samples showed significant differences in numbers of $\underline{H}$. azteca among the three shores during the breeding season. In 1981, a majority of samples showed no significant difference in total numbers of amphipods, or numbers of adults or juveniles among the three shores.

## Length-Weight

Statistical tests showed no significant difference between the weights of males and females, nor between preserved and unpreserved animals of the same size class. There were no


FIGURE 4: Relationship of the natural logarithms of weight (micrograms) to total length (millimeters) for Hyalella azteca in Dock Lake. $\quad(\mathrm{n}=588)$
$\begin{aligned} & \text { TABLE 1: Regression equations and correlation coefficients for selected morphometrics } \\ & \text { and length-weight of Hyalella azteca in Dock Lake, Ontario. The variables are } \\ & \text { as follows; } W \text { is dry weight in micrograms, TL is total length in mm, HL is head } \\ & \text { length in } \mathrm{mm} \times 10^{-2} \text {, A is antennal segment count (the ageing criterion). }\end{aligned}$
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seasonal weight differences within size classes, except for the April sample, in which the amphipods were significantly lighter than those of the same size classes from the June, August and October samples. On the assumption that these animals were lighter than the others due to depletion of body weight during winter, the April sample was omitted from the length-weight regression, leaving 588 weight data.

Figure 4 shows the relationship of weight in micrograms to total length in millimeters (see also Appendix A: 1 and A: 2). The regression equation is included in Table 1.

Total length and head length were judged effectively equivalent in accuracy (see Table l), and total length chosen for subsequent analysis, as the more convenient measurement.

Mean egg weight was $5.4 \pm 0.1$ micrograms (920 eggs, 9 weight measurements).

Characteristics of the Population

Population Density
Figure 5 shows the density of Hyalella azteca on the three shores over the ice-free seasons of 1980 and 1981. On the Carex shore, peak population numbers occurred on August 6, 1980 (17,459 animals $/ \mathrm{m}^{2}$ ), and July 29, 1981 (10,579 animals/m ${ }^{2}$.

Growth
Using the instar specific antennal segment counts from
Cooper (1965) and Geisler (1944) to age specimens, average head


FIGURE 5: Mean population densities of Hyalella azteca in three habitat types of Dock Lake (Carex bed, Typha stand and Gravel beach), for the ice-free seasons of 1980 and 1981.
lengths and total lengths for instars 2 to 9 were calculated from the 1980 production data and the length-weight data (Table 2). Up to instar 7, the head lengths agreed closely with Cooper's data. Head length and total length plotted against antennal segment count, as well as the length-weight data, display a great deal of scatter in the adult sizes (see Appendix A: 1).

Growth is a function of temperature (Figure 6). Molt rates (Cooper, 1965; Geisler, 1944) graphed against the weight of each instar show that growth is much slower at $15^{\circ} \mathrm{C}$ than at $20^{\circ} \mathrm{C}$.

All free-swimming young from the field, or observed in the laboratory were instar 2 or older, and the majority of those taken from brood pouches were instar 2 (13 antennal segments; Cooper, 1965). No evidence of molting within the brood pouch was found.

Table 2 was used to construct size classes for the size frequency analysis. From instars 3 to 9, total lengths increase in approximately 0.5 millimeter increments. The largest individuals caught were 7.7 mm . Accordingly, thirteen size classes with midlengths from 1.5 to 7.5 mm were constructed. Size classes 1 to 7 correspond to instars 3 to 9. Instars 2 and 3 (free-swimming sizes) were combined into size class 1.

Age Composition
The size frequency histograms (Appendix B) show that three
Mean head lengths and total lengths for instars 2 to 9 of Hyalella aztec in Dock Lake, $n$ : number of animals of each instar used to calculate mean lengths; $s$ : standard deviation.

 $=1$ 어 으 in $\infty$ 으 i in




 | Av. Antenna |
| :--- |
| Segment No. |

$\cdots \pm \underset{\sim}{n} \underset{\sim}{\dot{N}} \dot{\sim} \dot{n}$



FIGURE 6: Growth curve for Hyalella azteca, from instars 1 through 10. Molt rates for juveniles, and incubation rates (corresponding to adult female molt rates) are taken from Cooper (1965). Weight is weight at average length for each instar, calculated from the length-weight regression.
major hatch peaks occurred in both 1980 and 1981. Because May 1981 was colder, the first hatch occurred several days later than in 1980. By the third hatch of either year, animals from the first brood had grown, into the adult size classes, and were ovigerous. Comparison of the modal progression of hatches on the histograms, instar analysis data (Cooper, 1965), and molt rate calculations all confirm that the first hatch grows into and merges with its parents, making the separation of year classes difficult. Within a year class, each hatch eventually merges indistinguishably into a "pool" of adults (see Appendix B).

In Table 3, the first hatch of 1981 can be followed from its appearance until the last sample of the season. Molt rate calculations are compared to the locations of the hatch peak in each size frequency histogram (Appendix B). This first brood of 1981 first appeared in sample 4, occupying the first three size classes. Molt rate calculations and the histograms for the Typha shore indicate that its remnants are still present in sample 10 (mid-October).

The first hatch of the 1980 year class occurred on June 3-4, 1980. The last appearance of overwintered adults from this cohort was on July 29, 1981; a total of 412 days. The average life span of an individual is about 365 days, however there is considerable individual variation.

Reproduction
Gravid females were captured in Dock Lake from May 23 to
TABLE 3:

| $\begin{array}{l}\text { approximate } \\ \text { size classes } \\ \text { occupied by } \\ \text { hatch (size } \\ \text { frequency } \\ \text { histograms) }\end{array}$ |
| :--- |
| $1,2,3$ |
| $3,4,5$ |
| $5,6,7,8$ |
| $7,8,9$ |
| $8,9,10$ |
| $9,10,11$ |
| 9,10 |
| $10,11,12,13$ | əZ!s | $\begin{array}{c}\text { number } \\ \text { of adult } \\ \text { molts } \\ \text { per } \\ \text { interval }\end{array}$ | $\begin{array}{c}\text { calculated } \\ \text { size } \\ \text { classes } \\ \text { occupied } \\ \text { by hatch }\end{array}$ |
| :---: | :---: |
| 0.7 | $\frac{1,2,3^{*}}{}$ |
| 1.2 | $3,4,5$ |
| 1.3 | $6,7,8$ |
| 1.3 | $7,8,9$ |
| 1.2 | $8,9,10$ |
| 0.8 | $9,10,11$ |
| $(1.3)$ | $10,11,12$ |
|  | $11,12,13$ |


 mean duration of Cooper (1965)

| juvenile |
| :---: |
| instars |
| at mean |
| temperature |



|  |  | $\bar{\sim}$ |  |  |  | $\cdots$ | $\begin{aligned} & \Omega \\ & \vdots \end{aligned}$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | $\pm$ | $\pm$ | $\pm$ | $\pm$ |  |  | ¢ |



[^0]August 26 in 1980, and from May 20 to August 12 in 1981. First age to maturity was 24 days in 1980, and 30 days in 1981. The first hatch that matured, bred in their first summer in both years; however, reproduction by the smaller adults of the second brood ceased much earlier than among the larger ones (of the first hatch). The third hatch failed to breed during its first summer.

Brood counts of gravid females correlated with animal size (see Appendix A: 3). Mean brood size ranged from 5 to 36 eggs for female total lengths of 3.8 and 7.0 mm . respectively. The largest brood found in the subsampled animals was 42 eggs, in the marsupium of a 6.7 mm . female. There was considerable variation in brood size, particularly among the larger sized animals. Among the three shores, brood counts for any given size class tended to be highest on the Carex shore, slightly lower on the Gravel shore, and lowest on the Typha shore; however, these trends were not statistically significant.

## Production

Waters (1977, 1979, 1981) indicates that use of the largest size class of a benthic organism may lead to overestimates of production, therefore the "times loss" factor in all size frequency estimates was arbitrarily reduced (as suggested by Waters and Crawford, 1973) from 13 to 12 . The mean annual number of animals in each size class (from the computer program output) was multiplied by the mean weight of an individual of that size class, to compare
relative contributions to biomass. Size class 13 contributed less than size class 1 , which supports the above decision. Similarly, instars 1 and 2 are not part of the free-swimming (and "catchable") population, therefore the first size class (mean length 1.5 mm ) was used as the weight at hatching, since this is the length at which H. azteca leaves the marsupium.

Cohort $G$ is calculated as the natural log of mean weight of the largest size divided by the mean weight at hatching. The mean weights of size classes 1 and 12 being 15 and 1487 micrograms, cohort G was therefore 4.596. This value was divided by the growing season (Waters, 1981); delimited by the length of time in which water temperatures are $10^{\circ} \mathrm{C}$ or higher ( 150 days in 1980 plus 40 days to the disappearance of the cohort in 1981, for a cohort growth season of 190 days). The daily $G$ was thus 0.024192 .

Annual production was calculated for 1980 and 1981 using the size frequency and cohort $G$ methods, for the three shores (Table 4 , and Appendix C). To obtain a full year of samples as required by the size frequency method, the first sample of 1981 was used to represent the last sample of 1980. This gave a sampling year from ice-out 1980 to ice-out 1981. Since the fall 1980 and spring 1981 samples were almost identical, winter mortality was assumed to be minimal; and the fall 1981 sample was therefore used as an approximation of the April 1982 population, for the 1981 calculation. Production estimates for the littoral zone of Dock Lake were 1.24 and $1.40 \mathrm{~g} / \mathrm{m}^{2}$, for 1980 and 1981 respectively.
methods,
Annual
that shore
Annual dry weight production $\left(\mathrm{g} / \mathrm{m}^{2}\right)$, by the size frequency and cohort $G$
biomass and $\mathrm{P} / \mathrm{B}$ ratios; for 1980 and 1981 on three shores of Dock Lake.
means are calculated by weighting each shore value by the proportion of
type in the lake (Carex $34 \%$, Typha $42 \%$, Gravel $24 \%$ ).
TABLE 4:
$P / \bar{B}$
$\begin{array}{r}4.096 \\ 3.676 \\ 4.004 \\ \hline 3.933\end{array}$



METHODS
size
frequency


| 0 | $\hat{0}$ | $i n$ | 0 |
| :---: | :---: | :---: | :---: |
| 0 | 0 | 0 |  |
| $\dot{0}$ | 0 |  |  |
| $\dot{m}$ | - | $\dot{0}$ | - |

0

Cohort production was estimated for the 1980 year class using the size frequency (Table 5), daily $G$, cohort $G$, instantaneous growth, removal summation and Allen curve methods. These seven methods gave comparable estimates of annual production, ranging from 1.07 (instantaneous growth) to 1.46 (cohort G) $\mathrm{g} / \mathrm{m}^{2}$ (Table 6). Calculation of instantaneous growth, removal summation, Allen curve, and daily growth are given in Appendix $D$.

Annual $\mathrm{P} / \overline{\mathrm{B}}$ ratios on the three shores were: Carex 4.0 ,
Typha 3.6, and Gravel 4.1 for the 1980 cohort. The weighted annual $P / \bar{B}$ ratio was 3.9. By the size frequency method, annual production of this year class was $1.23 \mathrm{~g} / \mathrm{m}^{2}$ (Table 6).
lations for Hyalella azteca on three shores
has been reduced from 13 to 12 to avoid over-
ze class.

| WEIGHT AT | TOTAL WEIGHT | PRODUCTION |
| :--- | :--- | :--- |
| LOSS | AT LOSS | $\left(G / M^{2} / Y E A R\right)$ |

PRODUCTION
(G/M²/YEAR)
PRODUCTION
(G/M2/YEAR)

## $\underset{\sim}{N}$

TOTAL WEIGHT
AT LOSS

 -0.031028
0.113419
 0.035544
0.005306


 WEIGHT AT
LOSS
0.000023
0.000049
0.000091
0.000150
0.000229
0.000330
0.000460
0.000628
0.000825
0.001058
0.001331
0.001648
0.001827

$$
\begin{gathered}
\times \\
\dot{N} \\
\pm \\
\stackrel{1}{ \pm} \\
\times
\end{gathered}
$$

MEAN WEIGHT
(G/INDIVIDUAL)
0.000015
0.000035
0.000069
0.000119
0.000188
0.000280
0.000389
0.000545
0.000724
0.000939
0.001192
0.001487
0.001827
12.

PRODUCTION
(G/M2/YEAR)
0.999



| TABLE 6: | Biomass, and production $\left(\mathrm{g} / \mathrm{m}^{2}\right)$ using 7 methods, for the 1980 year class of Hyalella azteca on three shores in Dock Lake, Ontario. Allen curves 1 and 2 are calculated from hand smoothed curves, and data connected by straight lines, respectively. Cohort $G$ is 4.596 and daily $G$ is 0.024192 . Cohort values are calculated over a 412 day period (presence of the 1980 year class in the environment) and annual values are cohort figures $\times 365 / 412$. |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | BIOMAS |  |  | METHODS |  |  |  |  |
|  |  | size <br> frequency | instantaneous growth | $\begin{gathered} \text { Cohort } \\ G \end{gathered}$ | $\underset{G}{\text { daily }}$ | removal summation | Allen Curve 1 | Allen Curve 2 |
| COHORT |  |  |  |  |  |  |  |  |
| Carex | 0.468 | 1.887 | 2.533 | 2.153 | 2.979 | 2.557 | 2.579 | 2.171 |
| Typha | 0.335 | 1.212 | 0.744 | 1.541 | 0.931 | 0.745 | 0.862 | 0.746 |
| Gravel | 0.245 | 0.999 | 0.138 | 1.127 | 0.651 | 0.124 | 0.551 | 0.378 |
| weighted mean | 0.359 | 1.391 | 1.206 | 1.650 | 1.560 | 1.212 | 1.371 | 1.142 |
| Annual |  |  |  |  |  |  |  |  |
| Carex | 0.415 | 1.672 | 2.244 | 1.907 | 2.634 | 2.265 | 2.285 | 2.284 |
| Typha | 0.297 | 1.074 | 0.659 | 1.365 | 0.825 | 0.660 | 0.764 | 0.659 |
| Grave I | 0.217 | 0.885 | 0.122 | 0.998 | 0.577 | 0.110 | 0.488 | 0.296 |
| weighted mean | 0.318 | 1.231 | 1.069 | 1.462 | 1.382 | 1.074 | 1.215 | 1.124 |

## Distribution

Hyalella azteca is rarely found below 2 to 3 meters (Hargrave, 1970). For example, higher densities of this amphipod occurred in the shallower areas of both West Blue Lake, Manitoba (Biette, 1969), and Marion Lake, British Columbia (Mathias, 1971). A similar distribution occurred in Dock Lake. Shallow water seems to be the preferred habitat for this amphipod.

Habitat preference and depth distribution relates to environmental heterogeneity. Macrophyte beds offer refugia from predation for both young and adult macrobenthos, as well as increased surface area for periphyton growth (Hall, Cooper and Werner, 1970). Among the three shore habitats of Dock Lake, the gravel substrate provided the least structural complexity, and the Carex beds the greatest, while Typha was intermediate. The substrate between stalks of Typha was a flocculent ooze quite different from the structured litter of the Carex beds. These habitats thus form a gradient of shelter and food.

In Dock Lake $H$, azteca does not occur below 1.5 meters (Figure 3). This corresponds to the depth limit of macrophytic growth. Below this depth, only soft lake sediments are found. Mathias (1971) states that $\underline{H}$. azteca displays poor growth when
fed on such lake sediments, but does grow well on a diet of periphyton. Light attenuation causes epibenthic periphyton production to decrease with depth (Hargrave, 1970). Below 1.5 meters in Dock Lake, there is little periphyton (preferred food) or substrate (macrophytes) available; thus $\underline{H}$. azteca is apparently limited to depths less than 1.5 meters by a combination of food and substrate.

Food may be the most important factor of habitat quality affecting distribution and population density of $\underline{H}$. azteca in any given body of water. Differences in density with habitat type (see Figure 5) suggest that both type and quantity of vegetation (food) are important to the distribution and standing crop of H. azteca. Growth and production of Asellus racovitzae and Gammarus fasciatus in the lab were "profoundly affected" by the type of algae available as food (Swiss and Johnson, 1976). In Gammarus pulex, weight increase and molt rate at a given temperature also reflect the varying nutritional values of different diets (Willoughby and Sutcliffe, 1976). Hargrave (1970) found that in ㅂ. azteca, the ingestion rate and assimilation efficiency were lower on a diet of blue-green algae than on green algae and diatoms. The latter are a major component of the periphyton, which encrust the surface of macrophytes as well as other submerged substrates (Wetzel, 1975). Gerking (1962) has commented on the causal relationship of variability in abundance and distribution of macrophytes to the spatial distribution of benthos. Further studies involving
surveys of available food in different habitats, and analysis of the diet of $\underline{H}$. azteca captured from these habitats, would test the hypothesis that food is the most important limiting factor.

## Life Cycle

Growth
The mean instar sizes for $\underset{H}{H}$ azteca listed in Table 2 are based on an entire growing season, and can be considered "annual" means. The slight differences between the two years possibly represents bias introduced by using different numbers of animals from different seasons within each calculation. Although the weight of animals within a size class was the same from month to month, variations in length at a given age, caused by the temperature dependent growth (Figure 6), may have been present.

Antennal segment count could not be used to accurately age adult $\underline{H}$. azteca, because head length and total length plotted against antennal segment count (Appendix A: l) showed great scatter as antennal segment count increased. Wilder (1940) and Geisler (1944) also disclaim the effectiveness of antennal segment number as an ageing criterion. The standard deviation of head and total length for each instar also increased with age (Table 2). This increased variability with age could be a function of the interaction between temperature control of growth (ie, molt) rates and of maximum size in older animals (see de March, 1978).

Juvenile instars were identified by antennal segment counts.

Very few young $\underline{H}$. azteca taken from marsupiae corresponded to the first instar (12 antennal segments) as reported by Cooper (1965). The first visible segment of antenna 2 , which is actually the third segment (Bousfield, 1973, p. 20), is recessed in newly hatched animals, and becomes more evident after several days. Wilder (1940) lists instar 1 as having 13 antennal segments and total length of 1.3 mm , which agrees with Table 2. All other Dock Lake instars match the Sugarloaf Lake data (Cooper, 1965). A carefully planned rearing experiment would resolve the discrepancy in antennal segment count.

Growth in H. azteca is approximately linear, for instars 1 through 10 (Figure 6). Higher water temperature results in a steeper growth curve due to the increased rate of growth. Similar temperature dependent growth (with no growth at $10^{\circ} \mathrm{C}$ ) has been demonstrated for crayfish by Pratten (1980). Growth rates, adult body size and fecundity of some aquatic insects are also dependent on temperature during development (Sweeney and Vannote, 1978).

In Dock Lake, the first brood of 1980 developed at 18 to $20^{\circ} \mathrm{C}$, after which the temperature dropped to approximately $16^{\circ} \mathrm{C}$ for a week (Figure 2). According to de March (1978), this temperature regime should produce medium sized adults ( 5 to 6 mm .). The first brood of 1981 developed at 15 to $18{ }^{\circ} \mathrm{C}$, which should give large adults ( 5.5 to 7 mm .). In the size frequency histograms (Appendix B), these growth patterns were evident as an accumulation of small
adults in 1980; and the presence of large adults in the last sample of 1981 , identified by molt rate calculations as belonging to the first brood. Temperature control of adult length made it impossible to separate the contribution from each brood to the pool of adults, since all adults do not grow to the same maximum size.

Temperature dependent growth rates may be modified by population density. Wilder (1940) has shown that, given the same temperature, lab populations of $\underline{H}$. azteca tend to produce somewhat smaller adults at high density than at low density. This could explain the very large individuals that appeared on the Typha and Gravel shores, but were not evident in Carex samples, in fall, 1981.

## Voltinism

Cooper (1965) interpreted the Sugarloaf Lake population as bivoltine, with separate summer and winter cohorts. The Marion Lake (Mathias, 1971) and West Blue Lake (Biette, 1969) populations were both univoltine. The Dock Lake population was interpreted as univoltine, based on the size frequency histograms and molt rate calculations. Temperature dependent mean size of adult instars (de March, 1978) contributes to the difficulty of interpreting these data. Waters (1981) also comments on the difficulty of stating exact life cycle length in an amphipod with continuous reproduction and high individual variability.

The voltinism of $\underline{H}$. azteca could change with climate. Butler (1982) describes a 7 year life cycle for two Alaskan species of Chironomus, and changes in voltinism with latitude and altitude are also known in the Odonata (Ingram and Jenner, 1976; Deacon, 1979). Also, a small proportion of the population could differ in voltinism from the rest. A population of the odonate Enallagma aspersum was found to be $8 \%$ bivoltine, $92 \%$ univoltine (Ingram and Jenner, 1976). The unavailability of an emergence curve makes it difficult to determine such a pattern from the size frequency data for $\underline{H}$. azteca, since animals of different ages merge into the same size classes. If a small portion of the Dock Lake population was bivoltine, or if the average life span was less than 365 days, the production estimates and $P / \bar{B}$ ratios in Tables 4 and 5 were slightly underestimated.

## Reproduction

In Dock Lake, individuals born early in the season bred during their first summer. The data from this study, and from West Blue Lake (Biette, 1969) suggest that $\underline{H}$. azteca may mature and breed in its first summer, and breed again in its second summer if it survives the winter. The similarity of early spring and late fall size frequency and density data (Appendix B, and Figure 5) supports the hypothesis that the overwintering population is very stable, with minimal mortality (Cooper, 1965).

$$
\text { Comparison of data from four studies of } \underline{H} \text { azteca (Table 7) }
$$

shows that the onset of reproduction, as indicated by the first
appearance of eggs in marsupiae, corresponds closely with $16^{\circ} \mathrm{C}$; the critical threshold temperature reported by de March (1977) for reproduction in this animal. The 12L-12D daylength which induces and terminates reproduction at this temperature begins in midMarch and ends in late September at the latitude of Dock Lake (Bracken, 1982).

If the latitudinal difference of approximately $10^{\circ}$ between Sugarloaf Lake and West Blue Lake can be assumed to make little difference in the date of 12L-12D daylength, then north temperate populations of $H$. azteca are prevented from early spring breeding only by the temperature threshold, since the photoperiod threshold is reached while the lakes are still ice covered. Termination of breeding may depend on slightly different thresholds. In all four studies, cessation of breeding occurred at about the same time as temperature dropped below $18^{\circ} \mathrm{C}$. At this point, daylength had decreased to less than 14 hours of light. An age-specific factor may be involved as well, since young animals that matured late in the season did not breed, while older animals were still ovigerous.

Hence, both temperature and photoperiod must be in the optimal ranges of over 12 hours of light and at least $16^{\circ} \mathrm{C}$ (de March, 1977) for reproduction to occur. In the tropics, where there is little variation in daylength, temperature and/or some other parameter such as food may control reproduction. Sixteen hours of daylight induces reproduction at temperatures below $16^{\circ} \mathrm{C}$ (de March, 1977).

This may be of crucial importance to far northern populations of H. azteca.

## Production

Production estimates for Hyalella azteca range from 1.2 to $1.9 \mathrm{~g} / \mathrm{m}^{2}$ (Table 7). Production estimates for larger littoral zone amphipods are higher, ranging from 2.9 (Marchant and Hynes, 1981) to a maximum of $27.1 \mathrm{~g} / \mathrm{m}^{2}$ (Waters and Hokenstrom, 1980) for Gammarus pseudolimnaeus, and from 3.8 (Iversen and Jessen, 1977) to $12.9 \mathrm{~g} / \mathrm{m}^{2}$ (Welton, 1979) for G. pulex. Differences in annual production within a species may be due more to differences in population densities and recruitment rates than to growth rates of individuals (Marchant and Hynes, 1981). The difference in population density among the three shores in Dock. Lake (Figure 5) is reflected in the variability in biomass, production and $P / \bar{B}$ ratios (Tables 4 and 6).

The $P / \bar{B}$ ratios listed in Table 7 suggest divergence between cohort $G$ and the $P / \bar{B}$ ratio for $\underline{H}$. azteca. The higher values for the cohort and daily $G$ methods as compared to the size frequency method in Tables 4 and 6 support this hypothesis. Therefore, the production estimate for West Blue Lake may be an overestimate, as the cohort $G$ of 4.6 from the Dock Lake data was used to calculate it. This was deemed applicable to the West Blue population because the size frequency histograms in Biette (1969) indicated a maximum
size similar to the Dock Lake population.
The close agreement between annual production and $P / \bar{B}$ ratios for 1980,1981 and the 1980 cohort values (Tables 4 and 6) supports the hypothesis that $H$. azteca is univoltine, since the annual and cohort $P / \bar{B}$ ratios are the same. In a bivoltine species, the annual $P / \bar{B}$ ratio is twice as high as the cohort value (Waters, 1979). In addition, this agreement further validates the accuracy and usefulness of the size frequency method in analyzing populations with overlapping cohorts.

Several error factors may have contributed to the greater production value for 1981 than 1980 , despite the warmer temperature regime of the latter year. Considerable biomass may have been lost in the sugar flotation of early 1980 Typha samples, which would contribute to low weighted mean production. In both years, the earliest samples were slightly lower than subsequent ones despite the fact that no reproduction had occurred. This suggests that in spring, there is a gradual onshore movement of animals. The second sample captured more animals than the first, and as a result, biomass was underestimated twice in 1980 (early spring 1980 and 1981; the estimates are calculated from ice-out to iceout) and only once in 1981 (since the spring 1982 "sample" is an approximation); thus the 1981 production estimate would be larger than the 1980 estimate.

These error factors are slight when compared to the
effects of sampling error. Standard errors of the mean numbers of adults and juveniles per sample usually ranged from one half to one and one half times the mean. Hence differences between 1980 and 1981 are probably attributable to sampling error, which is normally the largest error factor in any production estimate (Marchant and Hynes, 1981; Waters, 1979).

Temperature is an important factor in the life history of benthic invertebrates (see Hynes, 1970; Lehmkuhl, 1979), because it influences features such as growth, voltinism and length of aquatic life, which are all critical to secondary production (Waters, 1979). Hyalella azteca is very temperature sensitive (Geisler, 1944; Cooper, 1965; de March, 1978). Table 7 compares annual production, biomass, and $P / \bar{B}$ ratios of four populations of this amphipod. Figure 7 illustrates the relationship of temperature to these production and biomass data in two ways; the number of days that the water temperature of the habitat being sampled is above $10^{\circ} \mathrm{C}$ (D10) and $20^{\circ} \mathrm{C}$ (D20). The former is an interaction of climate and lake morphometry, and is a factor limiting the geographic range of $\underline{H}$. azteca (Bousfield, 1958). The latter temperature range measures the length of optimal growth conditions for this amphipod (Cooper, 1965; de March, 1978). Table 8 lists the regression equations and correlation coefficients for Figure 7.

In both Marion Lake and West Blue Lake, temperatures exceed $20^{\circ} \mathrm{C}$ during less than two weeks of the year. Production values of just over $1 \mathrm{~g} / \mathrm{m}^{2}$ have been reported for both lakes (Mathias,

$$
\begin{aligned}
& \text { TABLE 7: Comparison of dry mass annual production }(P) \text {, annual biomass }(\bar{B}) \text { in grams per square } \\
& \text { meter, and } P / \bar{B} \text { ratios for four populations of Hyalella azteca. The asterisk indicates } \\
& \text { an extrapolation from the wet weight mean annual biomass given in Biette (1969). This } \\
& \text { value was divided by } 6 \text { to convert to dry weight (Waters, } 1977 \text { ), then multiplied by the } \\
& \text { cohort } G(=P / \bar{B}) \text { of } 4.6 \text { obtained in this study. }
\end{aligned}
$$

$$
g / d
$$

$$
3.9
$$

$$
4.8
$$

$$
4.2
$$

$$
(4.6)
$$

| $P$ | $\bar{B}$ | $P / \bar{B}$ | Reference |
| :---: | :---: | :---: | :--- |
| 1.23 | 0.32 | 3.9 |  |
| 1.93 | 0.40 | 4.8 | Cooper (1965) <br> IN Waters (1977) |
| 1.15 | 0.27 | 4.2 | Mathias (1971) |
| $1.20 *$ | 0.26 | $(4.6)$ | Biette (1969)* |



FIGURE 7: Interaction of climate with production (circles) and biomass (triangles) in $\mathrm{g} / \mathrm{m}^{2}$ dry weight of Hyalella azteca in four temperate zone lakes. Number of days above $20^{\circ} \mathrm{C}$ reflects optimal growth conditions; number of days above $10^{\circ} \mathrm{C}$ is an interaction of climate and lake morphometry.
TABLE 8:

| REGRESSION |  |  |  |  |  |  | CORRELATION COEFFICIENTS |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Number | Variables |  | Equ | at |  |  | Pearson | Kendal | Spearman |
| - | P, DIO | $P=$ | -0.151 | + | 0.009 | D10 | 0.93 | 0.33 | 0.40 |
| 2 | P, D20 | $P=$ | 1.040 | $+$ | 0.006 | D20 | 0.87 | 0.67 | 0.80 |
| 3 | $\bar{B}, \mathrm{D} 10$ | $\bar{B}=$ | 0.048 | + | 0.002 | D10 | 0.93 | 0.67 | 0.80 |
| 4 | $\bar{B}, \mathrm{D} 20$ | $\bar{B}=$ | 0.246 | + | 0.001 | D20 | 0.98 | 1.00 | 1.00 |

1971; Biette, 1969); the intercept of equation $2(D 20=0)$, ie, no optimal growth time, predicts production of approximately $1 \mathrm{~g} / \mathrm{m}^{2}$. The intercept of equation 1 (the DlO regression) is just below zero. No growth of young occurs at $10^{\circ} \mathrm{C}$ (Cooper, 1965), supporting the hypothesis that $\underline{H}$. azteca cannot tolerate habitats with temperature regimes below $10^{\circ} \mathrm{C}$. The effect of temperature could modify voltinism; arctic individuals probably require several years to complete development (see Butler, 1982), whereas tropical individuals could be multivoltine.

Brylinsky (1980) states that although biomass may be the same in lakes with long and short growing seasons (ie, in low and high latitudes), the number of generations produced, and therefore the amount of production per unit of biomass, increases with the length of growing season. Figure 7 supports this concept, with Biomass remaining relatively constant in the four lakes, while production varies with season length. Biomass, however, seems strongly related to habitat quality (particularly the amount and type of food) within the lake. Mean annual biomass differed noticeably among the three shores in Dock Lake (Table 4), despite almost identical temperature regimes (Figure 2). In H. azteca, biomass is apparently temperature independent, while production is temperature dependent.
that recommended its use for comparison to the others. The size frequency method is applicable where cohorts cannot readily be distinguished (Waters, 1977). The removal summation method apparently gives the most accurate estimates for a variety of growth curves and sampling regimes (Cushman, Shugart, Hildebrand and Elwood, 1978). The cohort $G$ is the simplest, and provides a check on the $P / \bar{B}$ ratio (Waters, 1981). Daily $G$ is a graphical method, useful when field and/or lab growth measurements are not available, and can illustrate voltinism (see Hall, Waters and Cook, 1980). The instantaneous growth method, and its graphical equivalent, the Allen curve, are used for populations with identifiable cohorts (Waters, 1977; Benke and Waide, 1977). Gillespie and Benke (1979) recommend use of a hand-smoothed Allen curve when sampling error is large (as in this study), and show that removal summation and increment summation equations are equal to an Allen curve estimate made by connecting the data points with straight lines. The difference between the removal summation and straight-line Allen curve estimates in Table 6 are probably due to sampling variability and overlapping broods.

Waters (1981) justifies the use of the size frequency method as a standard in the study of $\underline{G}$. pseudolimnaeus. As H. azteca is also a univoltine benthic amphipod with continuous reproduction over the optimal growth season, these same criteria apply. In studies which use more than one method, the size frequency estimate
generally gives a slightly higher result than the Allen curve, instantaneous growth or removal summation methods (see Waters and Hokenstrom, 1930; Wildish and Peer, 1981). However, the cohort $G$ and daily $G$ methods compare with or even exceed size frequency estimates (see Waters, 1981; and Table 6). This may be due to deviations of cohort $G$ from the cohort $P / \bar{B}$ ratio, caused by variations in growth patterns and population densities (Waters, 1969).

In the comparison of cohort production by seven methods (Table 6), the lowest value is given by the instantaneous growth method. Cushman et al (1978) state that when the assumption of exponential growth is violated, production is underestimated by this method. Since the growth curve for $H$. azteca (Figure 6) is more or less linear, this may be the case in this study. The removal summation and size frequency methods assume linear growth, and may therefore be more accurate for this amphipod. The effect of prolonged recruitment is evident in Appendix D: 8; the Allen curves do not exhibit the expected smooth decline from high density/low mean weight to low density/high mean weight (Waters, 1969).

The extended presence of newly hatched animals in the population, and mortality of large adults before smaller ones of the same age, lower the mean weights calculated from the samples, and result in a great many negative values within the instantaneous growth and removal summation calculations (see Appendix D). The
size frequency method is not affected by this, since weights are calculated from the length-weight regression rather thar field data; however it is subject, as are all other methods, to the effect of sampling error on mean numbers. The variety of methods available, and their differing applications, suggest that if the data are available, several methods of production estimation should be used. For Hyalella azteca, the size frequency and removal summation methods are probably the most useful and accurate.

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## APPENDIX

Appendix A: 1 Relationship of antennal segment count to total length (top) and head length (bottom) for Hyalella azteca in Dock Lake. In both, $n=358$.

2 Relationship of head length ( $\mathrm{mm} \times 10^{-2}$ ) to total length (mm) of Hyalella azteca in Dock Lake.

3 Relationship of female total length (mm) to brood count of Hyalella azteca in Dock Lake.

Appendix B: 1 Size frequency histograms for Hyalella azteca on the Carex shore of Dock Lake in 1980 and 1981. Eleven of the 24 samples from 1980 are so arranged that the dates corresponded as closely as possible with the 1981 samples, for comparison. Vertical scale: size classes 1-13, horizontal scale: $1 \mathrm{~mm}=5 \%$.

2 Size frequency histograms for Hyalella azteca on the Typha shore of Dock Lake in 1980 and 1981. As in B: 1.

3 Size frequency histograms for Hyalella azteca on the Gravel shore of Dock Lake in 1980 and 1981. As in B: 1.

Appendix C: 1 Size frequency estimate of annual production for Hyalella azteca on the Carex shore of Dock Lake (1980). The times loss factor has been reduced from 13 to 12 to compensate for the minimal contribution by size class 13. These calculations are from May 1980 to May 1981.

## APPENDIX (Continued)

Appendix C: 2 Size frequency estimate of annual production for $H$. azteca on the Typha shore of Dock Lake (1980). As in C: 1.

3 Size frequency estimate of annual production for $\underline{H}$. azteca on the Gravel shore of Dock Lake (1980). As in C: 1.

4 Size frequency estimate of annual production for $\underline{H}$. azteca on the Carex shore of Dock Lake (1981). These calculations are from May 1981 to an estimated sample for May, 1982.

5 Size frequency estimate of annual production for $H$. azteca on the Typha shore of Dock Lake (1981). As in C: 4.

6 Size frequency estimate of annual production for $H$. azteca on the Gravel shore of Dock Lake (1981). As in C: 4.

Appendix D: 1 Calculation of production of Hyalella azteca on the Carex shore, Dock Lake, by the instantaneous growth method. $\mathrm{G}=$ instantaneous rate of growth, $\mathrm{B}=$ standing crop, < $B$ > = mean standing crop over interval, $P=$ production over interval between successive sampling dates. Values are rounded from the full number of decimal places carried in the original calculations.

2 Calculation of production of $H$. azteca on the Typha shore, Dock Lake, by the instantaneous growth method. Calculated as in Table D: 1.

## APPENDIX (Continued)

Appendix D: 3 Calculation of production of $\underline{H}$. azteca on the Gravel shore, Dock Lake, by the instantaneous growth method. Calculated as in Table $D: 1$.

4 Calculation of production of Hyalella azteca on the Carex shore, Dock Lake ( 1980 cohort) by the removal summation method. $B=$ standing crop for each sample, Weight at loss is the geometric mean of the mean weights of two consecutive samples.

5 Calculation of production of Hyalella azteca on the Typha shore, Dock Lake ( 1980 cohort) by the removal summation method. As in Appendix D: 4.

6 Calculation of production of Hyalella azteca on the Gravel shore, Dock Lake ( 1980 cohort) by the removal summation method, As in Appendix D: 4.

7 Cohort production for Hyalella azteca on three shores in Dock Lake, by the daily growth method.

8 Cohort production for Hyalella azteca on three shores in Dock Lake, by the Allen curve method. Each Allen curve was constructed two ways; a hand drawn smoothed line (solid line) and straight lines between data' (dotted line).



APPENDIX A: 1 Relationship of antennal segment count to total length (top) and head length (bottom) for Hyalella azteca in Dock Lake. In both, $n=358$.


APPENDIX A: 2 Relationship of head length $\left(\mathrm{mm} \times 10^{-2}\right.$ ) to total length (mm) of Hyalella azteca in Dock Lake.


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APPENDIX A: 3 Relationship of female total length (mm)
    to brood count of Hyalella azteca in
Dock Lake.
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APPENDIX B: 2 Size frequency histograms for Hyalella azteca on the Typha shore of Dock Lake in 1980 and 1981. As in B: 1.


APPENDIX B: 3 Size frequency histograms for Hyalella azteca on the Gravel shore of Dock Lake in 1980 and 1981. As in B: 1.
$\begin{array}{cc} \\ \text { TOTAL WEIGHT } & \begin{array}{l}\text { PRODUCTION } \\ \text { AT LOSS }\end{array} \\ \left(G / M^{2} / Y E A R\right)\end{array}$
"
$\begin{array}{cc} \\ \text { TOTAL WEIGHT } & \begin{array}{l}\text { PRODUCTION } \\ \text { AT LOSS }\end{array} \\ \left(G / M^{2} / Y E A R\right)\end{array}$ -0.000377
-0.002949

0.011769




N
$\frac{7}{4}$
0
0
$x$ Carex shore ${ }^{1}$


azteca on the Gravel shore of
(y $\left.\forall \exists \lambda / Z^{W} / 9\right)$
NOIIOnOOyd
$\stackrel{\infty}{\infty}$
$\stackrel{\infty}{\circ}$
$\dot{\circ}$
$\stackrel{\infty}{\infty} \underset{\infty}{\infty}$
II

-0.000096
-0.001801
0.000198


9
$\vdots$
$\vdots$
$\vdots$
$\vdots$

 | $\underset{\infty}{\infty}$ |  |
| :--- | :--- |
|  |  |
|  |  |


$\times$

shore of
sample for
PRODUCTION
(G/M2/YEAR)
O
$\stackrel{N}{0}$
$i$
he Carex

TOTAL WEIGHT
AT LOSS
APPENDIX C: $4 \begin{aligned} & \text { Size frequency estimate of annual production for } \frac{H}{} . \\ & \\ & \text { Dock Lake (1981). These calculations are from May } 198\end{aligned}$

NUMBER
LOST
195.255
-149.120
143.551
46.910
-79.932
-18.221
18.465
-70.828
246.416
-21.556
22.228
2.923
0.595

N

0
0
0
$\times$

II

MEAN WEIGHT
(G/INDIVIDUAL)
0.000023
0.000049
0.000091
0.000150
0.000229
0.000330

0.000628
$\begin{array}{ccc}n & \infty & \bar{n} \\ \infty & 0 & \bar{m} \\ 0 & 0 & \bar{\circ} \\ 0 & 0 & 0 \\ 0 & 0 & 0\end{array}$

| $\infty$ |
| :--- |
|  |
| $\vdots$ |
| 0 |
| 0 |

x
12.


$$
\begin{aligned}
& 0.000023 \\
& 0.000049
\end{aligned}
$$



$$
\begin{aligned}
& \text { WEIGHT AT } \\
& \text { LOSS }
\end{aligned}
$$

$$
\begin{aligned}
& 0.000091 \\
& 0.000150
\end{aligned}
$$

$$
\begin{aligned}
& 0.000229 \\
& 0.000330 \\
& 0.000460
\end{aligned}
$$

$\begin{array}{ccccc}\sim & \infty & \bar{n} & \infty & \infty \\ 0 & 0 & ल & \overrightarrow{1} & \underset{\sim}{0} \\ 0 & 0 & \overline{0} & 0 & 0 \\ 0 & 0 & 0 & 0 & 0 \\ 0 & 0 & 0 & 0 & 0\end{array}$


|  | 1 Calculation of production of Hyalella azteca on the Carex shore, Dock Lake, by the instantaneous growth method. G = instantaneous rate of growth, $\mathrm{B}=$ standing crop, $\langle\mathrm{B}>=$ mean standing crop over interval, $\mathrm{P}=$ production over interval between successive sampling dates. Values below are rounded from the full number of decimal places carried in the original calculations. |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| DATE | Av.No. $/ \mathrm{m}^{2}$ | $\stackrel{B}{\left(\mathrm{~g} / \mathrm{m}^{2}\right)}$ | Mean Weight (micrograms) | G | $\begin{gathered} \left.<B_{2}\right\rangle \\ \left(\mathrm{g} / \mathrm{m}^{2}\right) \end{gathered}$ | $\begin{gathered} \mathrm{P} \\ \left(\mathrm{~g} / \mathrm{m}^{2}\right) \end{gathered}$ |
| Jun 6/80 | 2089.0 | 0.0453 | 21.7 |  |  |  |
|  |  |  |  | 0.3172 | 0.0720 | 0.0228 |
| Jun 17 | 3311.5 | 0.0987 | 29.8 |  |  |  |
|  |  |  |  | 0.5701 | 0.1069 | 0.0610 |
| Jun 22 | 2183.2 | 0.1151 | 52.7 |  |  |  |
|  |  |  |  | 0.8492 | 0.3566 | 0.3029 |
| Jun 27 | 4854.4 | 0.5982 | 123.2 |  |  |  |
|  |  |  |  | -0.1908 | 0.4988 | -0.0952 |
| Jul 2 | 3923.0 | 0.3993 | 101.8 |  |  |  |
|  |  |  |  | 0.2013 | 0.3197 | 0.0644 |
| Jul 7 | 1927.4 | 0.2400 | 124.5 |  |  |  |
|  |  |  |  | -0.0596 | 0.4286 | -0.0255 |
| Jul 13 | 5261.5 | 0.6172 | 117.3 |  |  |  |
| Jul 17 | 3074.9 | 0.2649 | 86.1 | -0.3092 | 0.4410 | -0.1364 |
|  | 3074.9 | 0.2649 |  | 0.4250 | 0.3615 | 0.1536 |
| Jul 22 | 3479.4 | 0.4582 | 131.7 |  |  |  |
|  |  |  |  | -0.1184 | 0.4939 | -0.0585 |
| Jul 27 | 4528.8 | 0.5297 | 117.0 |  |  |  |
| Aug 1 | 8199.3 | 1.1944 | 145.7 | 0.2194 | 0.8621 | 0.1891 |
|  |  |  |  | 0.0715 | 1.9621 | 0.1403 |
| Aug 6 | 17438.8 | 2.7298 | 156.5 |  |  |  |
|  |  |  |  | 0.2215 | 1.9018 | 0.4212 |
| Aug 11 | 5498.0 | 1.0738 | 195.3 |  |  |  |
|  |  |  |  | -0.0113 | 1.2230 | -0.0138 |
| Aug 16 | 7106.5 | 1.3722 | 193.1 |  |  |  |
|  |  |  |  | 0.1701 | 1.6606 | 0.2825 |
| Aug 21 | 8514.9 | 1.9490 | 228.9 |  |  |  |
|  |  |  |  | 0.0854 | 1.6978 | 0.1450 |
| Aug 26 | 5803.0 | 1.4465 | 249.3 |  |  |  |
| Aug 31 | 1878.6 | 0.5673 | 302.0 | 0.1918 | 1.0069 | 0.1931 |
|  |  |  |  | 0.0386 | 0.7823 | 0.0302 |
| Sep 7 | 3177.0 | 0.9974 | 313.9 |  |  |  |
|  |  |  |  | 0.0668 | 0.9003 | 0.0601 |
| Sep 14 | 2393.8 | 0.8033 | 335.6 |  |  |  |
|  |  |  |  | 0.2486 | 0.6932 | 0.1723 |
| Sep 21 | 1355.0 | 0.5831 | 430.3 | 0.1928 | 0.4268 | 0.0823 |
| Oct 5 | 518.6 | 0.2706 | 521.8 |  | 0.3042 |  |
|  |  |  |  | 0.0786 |  | 0.0239 |
| May 6/81 | 598.4 | 0.3378 | 564.5 |  | 0.6934 |  |
|  |  |  |  | 0.2696 |  | 0.1869 |
| May 20 | 1419.1 | 1.0490 | 739.2 |  |  |  |
|  |  |  |  | 0.2824 | 0.8569 | 0.2420 |
| Jun 3 | 678.2 | 0.6649 | 980.4 |  |  |  |
| Jun 17 | 438.8 | 0.4484 | 1021.8 | 0.0414 | 0.5566 | 0.0230 |
|  |  |  |  | 0.1678 | 0.4258 | 0.0714 |
| Jul 1 | 333.6 | 0.4032 | 1280.5 |  |  |  |
|  |  |  |  | -0.0081 | 0.2106 | -0.0017 |
| Jul 15 | 15.1 | 0.0181 | 1198.7 |  |  |  |
| Jul 29 | 9.2 |  |  | -0.2394 | 0.0134 | -0.0032 |
|  |  |  |  | -0.2648 | 0.0043 | -0.0012 |
|  |  |  | 724.0* |  |  |  |

[^1]| DATE | Av.No. $/ \mathrm{m}^{2}$ | $\left(\mathrm{B}_{(\mathrm{g}}{ }^{2}\right)$ | Mean Weight (micrograms) | G | $\begin{aligned} & <B> \\ & \left(\mathrm{g} / \mathrm{m}^{2}\right) \end{aligned}$ | $\stackrel{P}{\left(\mathrm{~g} / \mathrm{m}^{2}\right)}$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Jun 6/80 | 18.1 | 0.0004 | 23.2 |  |  |  |
|  |  |  |  | -0.4314 | 0.0003 | -0.0001 |
| Jun 17 | 7.3 | 0.0001 | 15.1 |  |  |  |
| Jun 22 | 246.7 | 0.0132 | 53.7 | 1.2701 | 0.0067 | 0.0085 |
|  |  |  |  | 0.3077 | 0.0179 | 0.0055 |
| Jun 27 | 308.3 | 0.0225 | 73.0 |  |  |  |
| Jul 2 | 14.5 | 0.0028 | 193.8 | 0.9762 | 0.0127 | 0.0124 |
|  |  |  |  | -0.0256 | 0.0017 | -0.0000 |
| Jul 7 | 3.6 | 0.0007 | 188.9 |  |  |  |
|  |  |  |  | -0.3412 | 0.0232 | -0.0079 |
| Jul 13 | 340.9 | 0.0458 | 134.3 | 0.0551 | 0.0270 | 0.0015 |
| Jul 17 | 58.0 | 0.0082 | 141.9 |  |  |  |
|  |  |  |  | -0.1510 | 0.0183 | -0.0028 |
| Jul 22 | 232.2 | 0.0283 | 122.0 | -0.0952 | 0.0200 | -0.0019 |
| Jul 27 | 105.2 | 0.0117 | 110.9 |  |  |  |
|  |  |  |  | 0.3872 | 0.1768 | 0.0684 |
| Aug 1 | 2092.5 | 0.3419 | 163.4 | 0.2854 | 0.3120 | 0.0891 |
| Aug 6 | 1298.4 | 0.2822 | 217.4 |  |  |  |
| Aug 11 | 431.5 | 0.0884 | 204.9 | -0.0589 | 0.1853 | -0.0109 |
|  |  |  |  | 0.1408 | 0.1371 | 0.0193 |
| Aug 16 | 787.1 | 0.1857 | 235.9 |  |  |  |
| Aug 21 | 823.3 | 0.1957 | 237.8 | 0.0078 | 0.1907 | 0.0015 |
|  |  |  |  | 0.2082 | 0.2359 | 0.0491 |
| Aug 26 | 942.8 | 0.2760 | 292.8 |  |  |  |
| Aug 31 |  |  |  | 0.0697 | 0.5251 | 0.0366 |
|  |  |  |  | 0.3507 | 0.4808 | 0.1686 |
| Sep 7 | 420.7 | 0.1875 | 445.8 |  |  |  |
|  |  |  |  | -0.0902 | 0.6182 | -0.0558 |
| Sep 14 | 2575.1 | 1.0488 | 407.3 | 0.1024 | 0.9189 | 0.0941 |
| Sep 21 | 1748.6 | 0.7890 | 451.2 |  |  |  |
| Oct 5 | 1581.4 | 0.8640 | 546.4 | 0.1914 | 0.8265 | 0.1582 |
|  |  |  |  | -0.0114 | 0.4506 | -0.0051 |
| May 6/81 | 68.9 | 0.0372 | 540.2 |  |  |  |
| May 20 | 383.2 | 0.2994 | 781.2 | 0.3689 | 0.1683 | 0.0621 |
|  |  |  |  | 0.2179 | 0.2096 | 0.0457 |
| Jun 3 | 123.3 | 0.1198 | 971.4 |  |  |  |
| Jun 17 | 43.5 | 0.0477 | 1097.0 |  | 0.0837 | 0.0102 |
|  |  |  |  | -0.0354 | 0.0277 | -0.0010 |
| Jul 1 | 7.3 | 0.0077 | 1058.9 |  |  |  |
| Jul 15 | 12.8 | 0.0155 | 1209.4 | 0.1329 | 0.0116 | 0.0015 |
|  |  |  |  | -0.2610 | 0.0095 | -0.0025 |
| Jul 29 | 3.8 | 0.0035 | 931.6 |  |  |  |
|  |  |  | 939.0* | 0.0079 | 0.0018 | 0.0000 |
|  |  |  | Total | 1 Product | on | 0.7442 |
| *estimate | from lengt | ight re | ression |  |  |  |


| APPENDIX D: $3 \begin{aligned} & \text { Calculation of production of } \frac{H}{} \text {. }{ }^{\text {azteca }} \text { on the Gravel shore, } \\ & \text { Dock Lake, by the instantaneous } \\ & \text { as in Table } \mathrm{D}: ~ \mathrm{growth} \text { method. Calculated }\end{aligned}$ |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| DATE | Av.No. $/ \mathrm{m}^{2}$ | $\left(\stackrel{B}{\left.g / m^{2}\right)}\right.$ | Mean Weight (micrograms) | G | < B > | $\left(\stackrel{P}{\left.\mathrm{~g} / \mathrm{m}^{2}\right)}\right.$ |
| Jun 6/80 | 127.0 | 0.0027 | 20.9 | 0.8083 | 0.0017 | 0.0013 |
|  |  |  |  |  |  |  |
| Jun 17 | 14.5 | 0.0007 | 46.9 |  |  |  |
|  |  |  |  | 0.4520 | 0.0249 | 0.0113 |
| Jun 22 | 667.3 | 0.0492 | 73.7 |  |  |  |
|  |  |  |  | 0.5024 | 0.0712 | 0.0358 |
| Jun 27 | 765.2 | 0.0932 | 121.8 |  |  |  |
| Jul 2 | 110.9 | 0.0175 | 157.6 | 0.2577 | 0.0553 | 0.0143 |
|  |  |  |  | 0.3850 | 0.0684 | 0.0263 |
| Jul 7 | 515.0 | 0.1193 | 231.6 |  |  |  |
|  |  |  |  | 0.0868 | 0.1475 | 0.0128 |
| Jul 13 | 696.0 | 0.1758 | 252.6 |  |  |  |
|  |  |  |  | 0.2446 | 0.1628 | 0.0398 |
| Jul 17 | 464.2 | 0.1498 | 322.6 |  |  |  |
|  |  |  |  | -0.3594 | 0.1414 | -0.0508 |
| Jul 22 | 591.1 | 0.1331 | 225.2 |  |  |  |
|  |  |  |  | 0.1367 | 0.2337 | 0.0320 |
| Jul 27 | 1294.5 | 0.3342 | 258.2 |  |  |  |
|  |  |  |  | -0.3597 | 0.3248 | -0.1168 |
| Aug 1 | 1749.7 | 0.3153 | 180.2 |  |  |  |
|  |  |  |  | 0.2578 | 0.2388 | 0.0616 |
| Aug 6 | 696.4 | 0.1624 | 233.2 |  |  |  |
| Aug 11 | 1932.9 | 0.2863 | 148.1 | -0.4540 | 0.2243 | -0.1018 |
|  |  |  |  | 0.2969 | 0.4041 | 0.1200 |
| Aug 16 | 2618.5 | 0.5220 | 199.3 |  |  |  |
|  |  |  |  | -0.1756 | 0.3022 | -0.0531 |
| Aug 21 | 493.1 | 0.0824 | 167.2 | 0.0765 | 0.2006 | 0.0154 |
| Aug 26 | 1766.2 | 0.3188 | 180.5 |  |  |  |
|  |  |  |  | -0.0022 | 0.1934 | -0.0004 |
| Aug 31 | 377.2 | 0.0679 | 180.1 |  |  |  |
| Sep 7 |  | 0.2028 |  | 0.4438 | 0.1353 | 0.0601 |
|  |  |  |  | -0.0456 | 0.1218 | -0.0055 |
| Sep 14 | 152.4 | 0.0409 | 268.2 |  |  |  |
|  |  |  |  | 0.4838 | 0.1051 | 0.0508 |
| Sep 21 | 389.1 | 0.1693 | 435.1 |  |  |  |
|  |  |  |  | -0.2635 | 0.4241 | -0.1118 |
| Oct 5 | 2031.0 | 0.6790 | 334.3 |  |  |  |
| May 6/81 | 3.6 | 0.0014 | 391.7 | 0.1585 | 0.3402 | 0.0539 |
|  |  |  |  | 0.7507 | 0.0098 | 0.0073 |
| May 20 | 21.8 | 0.0181 | 829.8 |  |  |  |
|  |  |  |  | 0.1236 | 0.0090 | 0.0011 |
| Jun 3 | 0.0 | 0.0000 | 939.0* |  |  |  |
|  |  |  |  | 0.1086 | 0.1536 | 0.0167 |
| Jun 17 | 293.6 | 0.3073 | 1046.7 |  |  |  |
|  |  |  |  | 0.1108 | 0.2004 | 0.0222 |
| Jul 1 | 80.0 | 0.0936 | 1169.4 |  |  |  |
|  |  |  |  | -0.1541 | 0.0552 | -0.0085 |
| Jul 15 | 16.9 | 0.0169 | 1002.4 |  |  |  |
| Jul 29 | 29.0 | 0.0338 | 1166.2 | 0.1514 | 0.0254 | 0.0038 |
|  |  |  |  | 0.0219 | 0.0169 | 0.0004 |
|  |  |  | 1192.0* |  |  |  |
|  |  |  |  | 1 Product | = | 0.1380 |


| APPENDIX | $\begin{array}{lll} \mathrm{D}: \quad 4 & \mathrm{Ca} \\ & \text { sh } \\ & & \text { me } \\ & \text { thi } \\ & \text { sar } \end{array}$ | Calculation of production of Hyalella azteca on the Carex shore, Dock Lake ( 1980 cohort) by the removal summation method. $B=$ standing crop for each sample, Weight at loss is the geometric mean of the mean weights of two consecutive samples. |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| DATE | Av.No./m ${ }^{2}$ | $\stackrel{B}{\left(\mathrm{~g} / \mathrm{m}^{2}\right)}$ | Mean Weight (micrograms) | Number Lost (per m${ }^{2}$ ) | Weight at loss $\left(g \times 10^{-4}\right)$ | $\begin{aligned} & \text { Weight Loss } \\ & \left(\mathrm{g} / \mathrm{m}^{2}\right) \end{aligned}$ |
| Jun 6/80 | 2089.0 | 0.0453 | 21.7 |  |  |  |
|  |  |  |  | -1222.5 | 0.0254 | -0.0311 |
| Jun 17 | 3311.5 | 0.0987 | 29.8 |  |  |  |
|  |  |  |  | 1128.3 | 0.0396 | 0.0447 |
| Jun 22 | 2183.2 | 0.1151 | 52.7 |  |  |  |
|  |  |  |  | -2671.2 | 0.0806 | -0.2152 |
| Jun 27 | 4854.4 | 0.5982 | 123.2 |  |  |  |
| Jul 2 | 3923.0 | 0.3993 | 101.8 | 931.4 | 1.1199 | 0.1043 |
|  |  |  |  | 1995.6 | 1.1258 | 0.2247 |
| Jul 7 | 1927.4 | 0.2400 | 124.5 |  |  |  |
|  |  |  |  | -3334.1 | 1.2085 | -0.4029 |
| Jul 13 | 5261.5 | 0.6172 | 117.3 |  |  |  |
| Jul 17 | 3074.9 | 0.2649 | 86.1 | 2186.6 | 1.0050 | 0.2197 |
|  |  |  |  | -404.5 | 1.0649 | -0.0431 |
| Jul 22 | 3479.4 | 0.4582 | 131.7 |  |  |  |
|  |  |  |  | -1049.4 | 1.2413 | -0.1303 |
| Jul 27 | 4528.8 | 0.5297 | 117.0 |  |  |  |
| Aug 1 | 8199.3 | 1.1944 | 145.7 | -3670.5 | 1.3056 | -0.4792 |
|  |  |  |  | -9239.5 | 1.5100 | -1.3952 |
| Aug 6 | 17438.8 | 2.7298 | 156.5 |  |  |  |
| Aug 11 | 5498.0 | 1.0738 | 195.3 | 11940.8 | 1.7483 | 2.0876 |
|  |  |  |  | -1608.5 | 1.9420 | -0.3124 |
| Aug 16 | 7106.5 | 1.3722 | 193.1 |  |  |  |
| Aug 21 | 8514.9 | 1.9490 | 228.9 | -1408.4 | 2.1024 | -0.2961 |
|  |  |  |  | 2711.9 | 2.3888 | 0.6478 |
| Aug 26 | 5803.0 | 1.4465 | 249.3 |  |  |  |
|  |  |  |  | 3924.4 | 2.7439 | 1.0768 |
| Aug 31 | 1878.6 | 0.5673 | 302.0 |  |  |  |
|  |  |  |  | -1298.4 | 3.0789 | -0.3998 |
|  | 3177.0 | 0.9974 | 313.9 | 783.2 | 3.2457 | 0.2542 |
| Sep 14 | 2393.8 | 0.8033 | 335.6 |  |  |  |
|  |  |  |  | 1038.8 | 3.8001 | 0.3948 |
| Sep 21 | 1355.0 | 0.5831 | 430.3 | 836.4 | 4.7385 | 0.3963 |
| Oct 5 | 518.6 | 0.2706 | 521.8 |  |  | 0.396 |
|  |  |  |  | -79.8 | 5.4273 | -0.0433 |
| May 6/81 | 598.4 | 0.3378 | 564.5 |  |  |  |
| May 20 | 1419.1 | 1.0490 | 739.2 | -820.7 | 6.4597 | -0.5302 |
|  |  |  |  | 740.9 | 8.5130 | 0.6307 |
| Jun 3 | 678.2 | 0.6649 | 980.4 |  |  |  |
|  |  |  |  | 239.4 | 10.0088 | 0.2396 |
| Jun 17 | 438.8 | 0.4484 | 1021.8 |  |  |  |
|  |  |  |  | 105.2 | 11.1124 | 0.1169 |
| Jul 1 | 333.6 | 0.4032 | 1280.5 |  |  |  |
| Jul 15 | 15.1 | 0.0181 | 1198.7 | 318.5 | 12.0359 | 0.3833 |
|  |  |  |  | 5.9 | 10.6347 | 0.0063 |
| Jul 29 | 9.2 | 0.0087 | 943.5 |  |  |  |
|  |  |  | 724.0* | 9.2 | 8.2650 | 0.0076 |
| *estimat |  |  |  | Total Prod | uction | 2.5567 |






$\begin{aligned} \text { APPENDIX D: } 7 & \text { Cohort production for Hyalella azteca on } \\ & \text { three shores in Dock Lake, by the daily } \\ & \text { growth method. }\end{aligned}$




APPENDIX D: 8 Cohort production for Hyalella azteca on three shores in Dock Lake, by the Allen curve method. Each Allen curve was constructed two ways; a hand drawn smoothed line (solid line) and straight lines between data (dotted line).


[^0]:    b: data in days

[^1]:    * estimated from length-weight regression

